

AD _____

GRANT NUMBER DAMD17-98-1-8107

TITLE: Growth Inhibition of Breast Tumor Cells by Hypodense and Normodense Eosinophilic Cell Lines

PRINCIPAL INVESTIGATOR: Paulette M. Furbert-Harris, Ph.D.

CONTRACTING ORGANIZATION: Howard University
Washington, DC 20059

REPORT DATE: July 1999

TYPE OF REPORT: Annual

PREPARED FOR: Commanding General
U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

DTIC QUALITY IMPROVED 4

20001019 026

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE July 1999	3. REPORT TYPE AND DATES COVERED Annual (15 Jun 98 - 14 Jun 99)	
4. TITLE AND SUBTITLE Growth Inhibition of Breast Tumor Cells by Hypodense and Normodense Eosinophilic Cell Lines			5. FUNDING NUMBERS DAMD17-98-1-8107	
6. AUTHOR(S) Paulette M. Furbert-Harris, Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Howard University Washington, DC 20059			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited			12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 words) We have shown that cell lines developed from metrizamide density gradient fractions of peripheral blood hypodense (BJA.060.22) and hyperdense (BJA.060.24) eosinophils significantly inhibited MCF-7 colony formation in a dose-dependent fashion with as much as 82% inhibition occurring at the E:T ratio of 1000:1 (BJA.060.22) and 85% inhibition of MDA-MB-231 cells at the same E:T ratio. At 1000:1 E:T ratio, the hyperdense eosinophil cell line BJA.060.24 inhibited MCF-7 colony formation by half that of the hypodense line (44%) and MDA-MB-231 colony formation by 70%. This inhibition is partially due to IL-4 as anti-IL-4 significantly abrogated the inhibitory activity of 24 hr eosinophil culture supernatants. Furthermore BJA.060.22 also bound tightly to MCF-7 multicellular tumor spheroids (MTS) and transmission EM analysis revealed the presence of peripheral blood eosinophils in the core of the tumor spheroid and within its spaces. MTS in co-culture with eosinophilic cell lines offer an excellent model system to clearly define a role for eosinophils in cancer and their upregulation by cytokines and other regulatory agents, both at the cellular and molecular levels.				
14. SUBJECT TERMS Breast Cancer Eosinophils, Cytokines			15. NUMBER OF PAGES 45	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

____ Where copyrighted material is quoted, permission has been obtained to use such material.

____ Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

____ Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

NA In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

X For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

NA In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

NA In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

NA In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

Paula E. Furber Harris 7/26/99
PI - Signature Date

Table of Contents

	Page Number
Front Cover	1
Standard Form 298	2
<i>Foreword</i> Forward	3
Table of Contents	4
Introduction	5
Body	5
Key Research Accomplishments	7
Reportable Outcomes	7
Conclusion	8
References	9
Appendices	10

5. INTRODUCTION

Eosinophils are principally known for their role in anti-helminthic responses as well as their inflammatory activity in allergic and asthmatic conditions (1). When activated, eosinophils release an array of inflammatory mediators including cytokines (2), many of which are toxic to the surrounding tissues. Moreover eosinophils, once activated by cytokines/chemokines can produce them and hence establish an autocrine cycle there by creating a constitutive eosinophilic presence at inflammatory sites (3-5). Eosinophils have been found in breast cancer infiltrates (6), however their active role in cancer, has not been well studied. We hypothesize that infiltrating eosinophils become activated and release cytokines into tumor inflammatory milieu and that these cytokines along with other released mediators inhibit growth of the tumor. In this study we have utilized eosinophils from allergic and asthmatic individuals with mild to moderate eosinophilia and cell lines established from them to investigate MCF-7 and MDA-MB-231 breast tumor cell growth inhibition in eosinophi:tumor co-culture assays.

6. BODY:

Propagation of Cell Lines. To date all eight eosinophilic cell lines have been retrieved from storage at -160°C , cultured in RPMI medium supplemented with sodium pyruvate (1mM), non essential amino acids (1mM), penicillin/streptomycin (50 units/50ug/ml, respectively), gentamycin (50ug/ml) and 10% fetal bovine serum; and refrozen. At present we have data with four of the cell lines.

Growth Inhibition of MCF-7 and MDA-MB-231 tumor cells by Eosinophilic Cell Lines.

a) Monolayer. Tumor cells (both MCF-7 and MDA-MB-231) were seeded into T_{25} culture flasks at 3×10^5 cells/flask and incubated overnight, (16-24hr) at 37°C , 5% CO_2 . Eosinophils were added at effector to target (E:T) ratios of 5:1 and 43:1 (for eosinophil hypodense line SD.031.22) and 5:1 and 14:1 (for eosinophilic hyperdense line SD.031.24) and the flasks were incubated for 3 to 4 days. Both cell lines SD.031.22 and SD.031.24 markedly inhibited both MCF-7 (figure 1B, 1C, 1D and 1E, respectively); and MDA-MB-231 cells (figure 2B, 2C, 2D and 2E, respectively). Results were similar when a third cell line, BJA.060.22 was tested against both MCF-7 (Figure 3) and MDA-MB-231 (Figure 4) tumor cells at E:T ratios of 1:1, (B) 20:1, (C) and 40:1, (D). The effector cells were removed, the flasks washed with phosphate buffered saline then stained with hematoxylin and eosin. Data were captured by photomicrography. In figure 5, peripheral blood eosinophil subpopulations hypodense (5A) and hyperdense (5B) confirm preliminary data of eosinophilic destruction of tumor cell growth, at E:T ratios as low as 2:1, (5B, 5C).

b) Colony Formation. MCF-7 tumor cells were seeded into the wells of a 6-well tissue culture plate at 500, 250, 100 and 50 cells/well to establish cloning efficiency and optimum concentration for the inhibition assay. The plates were incubated at 37°C , 5% CO_2 for ten days, until the colonies were visible by the naked eye. The plates were harvested, washed with PBS then stained with hematoxylin and eosin, and counted manually. In figure 6, the percent cloning efficiency for the four cell concentrations (triplicate experiments) were 50, 52, and 56% (500 cells); 60, 64, and 72% (250 cells); 140, 98, and 78% (100 cells) and 35, 45, and 80% (50 cells). From these data we originally selected both 250 and 100 cells, based on the percent efficiency, however we presently seed only 100 cells because of the facility in counting the colonies manually. Figure 7 shows colony inhibition of

MCF-7 cells by peripheral blood hypodense and hyperdense (A and B) eosinophils respectively. At 100:1 E:T ratios, both eosinophil subpopulations inhibited MCF-7 tumor colony formation by 79% and 80%, respectively. Hypodense eosinophilic cell line BJA.060.22 (Figure 8A) inhibited MCF-7 colony formation in a dose dependent manner at effector to target ratios of 10:1, 100:1, and 1000:1 (29%, 66% and 82%, respectively). The hyperdense cell line (BJA.060.24) inhibited colony growth by 33-44% at all ratios (figure 8B). At 100:1 it required 10-fold more cells from the cell lines to inhibit colony growth by the same extent as did the fresh eosinophils. Against MDA-MB-231 tumor cells BJA.060.24 eosinophilic cell line (figure 9B) inhibited colony formation dose dependently (3%, 43% and 70% at 10:1, 100:1, and 1000:1, respectively), while at 1000:1 BJA.060.22 (Figure 9A) dramatically inhibited colony formation by 93%.

Cytokine Presence in 24-hr Eosinophil Supernatants. 24hr supernatants from peripheral blood eosinophil hypodense and hyperdense fractions 22 and 24 were evaluated by enzyme-linked immunoassay (ELISA) analysis using commercial kits. Interleukin-4 and TNF α were present in varying levels in all six individuals tested. (Table 1). IL-4 concentrations ranged from 0 to > 1000 pg/ml while TNF α concentrations, which were far less than IL-4, ranged from 10-224 pg/ml. GM-CSF was only found in donor 6 at 450 pg/ml, and IL-5 was absent in all 6 samples.

24 hr Supernatants Inhibit MCF-7 Colony Formation. Based on the above results we evaluated 24hr supernatants for inhibition of tumor colony formation (Figure 10). Hypodense eosinophil fractions 22 inhibited MCF-7 colony formation by 30-50%. This inhibition was abrogated in all six of the samples when anti-IL-4 antibody was added to the tumor cells along with the supernatants. Supernatants from hyperdense eosinophil fractions (24) inhibited MCF-7 colony growth by 26-48%, and anti-IL-4 antibody abrogated the activity in 4/6 of the samples.

Infiltration of MCF-7 Multicellular Tumor Spheroids (MTS) by Peripheral Blood Hypodense and Hyperdense Eosinophil Fractions and by Eosinophil Cell Line.

a. MTS Production. MCF-7 multicellular tumor spheroids were developed by slightly modifying the method of Yuhas et al (7). Briefly, subconfluent monolayer cultures which were maintained (at 100% relative humidity, 95% air, 5% CO₂ at 37°C) in 10% RPMI complete medium. After trypsinization and cell count, the cells were dispensed into T₂₅ non-vented flasks (1 \times 10⁶ cells/flask) and rocked at 30 rpm at 37°C for 24- 48hr, after which time the spheroids were transferred to 100 mm petri dishes containing an overlayer of 0.3% noble agar in 10% RPMI medium. The dishes were then incubated at 37°C in a 5% CO₂ atmosphere for 7-14 days with regular feeding to study the growth characteristics of the MTS.

b. Eosinophil: MTS Co-culture Assay. As early as 48hrs, thirty MTS were transferred to a fresh petri dish with agar overlay. Eosinophils were added at ratios of 10:1 and 100:1 eosinophil: MTS. The dishes were incubated for 7 days, then observed microscopically for eosinophil attachment. MTS were also fixed in 4% glutaraldehyde and processed with en bloc uranyl acetate for electron microscopic analyses.

Fig (11) shows photomicrographs of MTS in culture for 7 days. They range in size from 100 μ m to 500 μ m. Characteristic necrotic cores can be seen in A and C. Photomicrographs in figure (12) show fresh eosinophils, hypodense (A,C and E) and hyperdense fractions (B,D and F) bound to spheroids, while in fig (13) eosinophil cell line BJA.060.22 is seen bound to the tumor spheroid. Transmission EM (Figure 14) analysis revealed eosinophils inside the MTS, both just inside the surface area and in the core. (Abstract to AACR)

7. Key Research Accomplishments

- ▶ Retrieval of all 8 eosinophilic cell lines
- ▶ Demonstration of functional cytotoxic/cytostatic activity with 4 of the lines
- ▶ Establishment of Multicellular Tumor Spheroids (MTS)
- ▶ Infiltration of MTS by eosinophils.

8. Reportable Outcomes

Hypodense and hyperdense eosinophils infiltrate MCF-7 breast multicellular tumor spheroids. Furbert-Harris PM, Harris D, Vaughn T, Parish-Gause D, Dunston GM, Abdelnaby A, Laniyan I, and Oredipe O. Proc Am Assoc Cancer Res. 40:455, 1999.

Furbert-Harris PM, Anderson D, Parish-Gause D, Vaughn T, Brown R, Laniyan I, Dunston GM, Abdelnaby A and Oredipe O.. Eosinophilic destruction of breast tumor cells in vitro is mediated by Interleukin-4. FASEB 13(4):A612, 1999.

9. Discussion/Conclusion

We hypothesized that activated eosinophils inhibit breast cancer cell growth by binding to the tumor cell, infiltrating the tumor and releasing inflammatory mediators, which include cytokines and these cytokines are known to have anti-cancer activity. The data presented have shown that subpopulations of hypodense and hyperdense activated eosinophils from individuals with mild to moderate eosinophilia inhibit the growth of MCF-7, and MDA-MB-231 breast tumor cells in vitro. Inhibition of colony formation was demonstrated with both activated eosinophils from peripheral blood as well as eosinophilic cell lines established from these hypodense and hyperdense subpopulations. At the effector to target ratio of 100:1, it required 10-fold more cells from the cell line to exact the same percent colony inhibition. This suggests that the fresh eosinophils are more activated than the cell lines and that perhaps treatment of these cell lines with cytokines will upregulate their level of activity. This is indeed one of the objectives of the study. Hence studies will be performed to treat cell lines with cytokines and other agents known to activate eosinophils. (e.g. IL-3, IL-4, IL-5, Ca ionophore), and evaluate their functional activity. The data indicating the absence of IL-5 and GM-CSF in 24hr culture supernatants of activated eosinophils is not 100% credible. Perhaps this is due to technical problems with ELISA Kits, particularly at the wash step. We are also still trying to select a single vendor for the ELISA Kits. There could also have been a problem with the kits themselves. We will introduce RT-PCR analysis for cytokine mRNA levels to complement the ELISA studies.

When eosinophils (both fresh and cell line) were co-cultured with MCF-7 MTS, (a model of micrometastatic nodules), the cells bound to the surface of the spheroid. This was seen by light microscopy: and transmission electron microscopy revealed the presence of eosinophils (fresh) within the spheroid. We will include scanning EM will include EM analysis of cell line infiltrated spheroids and also immunohistochemical analysis of the spheroids for the presence of cytokines and other eosinophil mediators (e.g. granular proteins). Moreover kinetic studies of eosinophil attachment and infiltration are needed.

The resistant line MBA-MB.231 does not grow very well under our culture conditions. Colony formation was difficult to obtain. Hence the data with this cell line may be equivocal. We therefore will select another resistant breast cell line for future studies.

These studies thus far, confirm the preliminary observations of eosinophil destruction of tumor cells in vitro. The multicellular tumor spheroid model, along with the eosinophil cell lines (we have further purified the cells by FACS sorting and are presently carrying out phenotypic analysis - manuscript in preparation) offer an exquisite model system for defining the role for eosinophils as anti-cancer effector agents and their upregulation by cytokines and other agents.

10. References

1. Gaga M, Frew AJ, Veronica AV and Kay AB. 1991. Eosinophil activation and T lymphocyte infiltration in allergen-induced late phase skin reactions and classical delayed type hypersensitivity. *J. Immunol* 147:816.
2. Obiri NI, Siegel JP, Varricchio F and Puri RK. 1994. Expression of high affinity IL-4 receptors on human melanoma, ovarian and breast carcinoma cells. *Clin Exp Immunol* 95(1):148.
3. Tepper RI, Coffman RL and Leder P. 1992. An eosinophil-dependent mechanism for the anti-tumor effect of interleukin-4 *Science* 257:548.
4. Nakajima H, Gleich GJ and Kita H. 1996. Constitutive production of IL-4 and IL-10 and production of IL-8 by normal peripheral blood eosinophils. *J. Immunol* 156(12):4859.
5. Elovic AE, Ohyama H, Sauty A, McBride J, Tsuji T, Nagari M, Weller PF, Wong DT. 1998. IL-4 dependent regulations of TGF-alpha and TGF-beta 1 expression in human eosinophils. *J. Immunol* 160(12):6121.
6. Samoszuk MK, Nguyen V, Gluzman I, Pham JH. 1996. Occult deposition of eosinophil peroxidase in a subset of human breast carcinoma. *Am J Pathol* 148(3):701-6.
7. Yuhas JM, Li AP, Martinez O, Ledman AJ. 1997. A simplified method for production and growth of multicellular tumor spheroids. *Cancer Res* 57:3639.

11. Apendices.

- Figure 1. Inhibition of MCF-7 Tumor Cell Growth by Eosinophilc Cell Lines
- Figure 1. Legend
- Figure 2. Inhibition of MDA-MB-231 Tumor Cell Growth by Eosinophilic Cell Lines
- Figure 2. Legend
- Figure 3. Inhibition of MCF-7 Tumor Cell Growth by Eosinophilic Cell Lines
- Figure 3. Legend
- Figure 4. Inhibition of MDA-MB-231 Tumor Cell by Eosinophilic Cell Line
- Figure 4. Legend
- Figure 5. Inhibition of MCF-7 Tumor Cell Growth by Peripheral Blood Eosinophils
- Figure 5. Legend
- Figure 6. Plating Efficiency of MCF-7 Tumor Cells
- Figure 6. Legend
- Figure 7. Growth Inhibition of MCF-7 Tumor Cell Colony Formation by Peripheral Blood Hypodense and Hyperdense Eosinophils
- Figure 7. Legend
- Figure 8. Growth Inhibition of Colony Formation by Eosinophilic Cell Lines
- Figure 8. Legend
- Figure 9. Inhibition of MDA-MB-231 Tumor Cell Colony Formation by Eosinophilic Cell Lines
- Figure 9. Legend
- Table 1. Cytokine Concentrations in 24hr Eosinophil Culture Supernatants (pg/ml)
- Table 1. Legend

Figure 10. Inhibition of MCF-7 Colony Formation by 24-hr Eosinophil Supernatants

Figure 10. Legend

Figure 11. MCF-7 Multi-cellular Tumor Spheroids

Figure 11. Legend

Figure 12. Peripheral Blood Hypodense and Hyperdense Eosinophils Bind to MCF-7 Multi-cellular Tumor Spheroids

Figure 12. Legend

Figure 13. Hypodense Eosinophilic Cell Line Binds to MCF-7 Multi-cellular Tumor Spheroids

Figure 13. Legend

Figure 14. Eosinophil Infiltration of MCF-7 Multi-cellular Tumor Spheroid

Figure 14. Legend

Abstract to AACR '99

Abstract to FASEB '99

**Fig. 1. Inhibition of MCF-7 Tumor Cell Growth
by Eosinophil Cell Lines**



Fig. 1. MCF-7 tumor cells were seeded into T₂₅ flasks at 3×10^5 cells/flask and allowed to grow to confluence (4-6 days) in media alone (A) or on the presence of hypodense eosinophilic cell line SD.031.22 at E:T ratios 5:1 and 43:1 (B and C) and hyperdense - (D and E) cell line SD.031.24 at E:T ratios 5:1 and 14:1.

Fig. 2. Inhibition of MDA-MB-231 Tumor Cell Growth by Eosinophil Cell Line



Fig. 2. MDA-MB-231 tumor cells were seeded similarly to MCF-7 in media (A) and with SD.031.22 (B) and SD. 031.24 (C) at similar E:T ratios 5:1, 43:1 and 5:1, 14:1, respectively.

Fig. 3. Inhibition of MCF-7 Tumor Cell Growth by Eosinophil Cell Lines

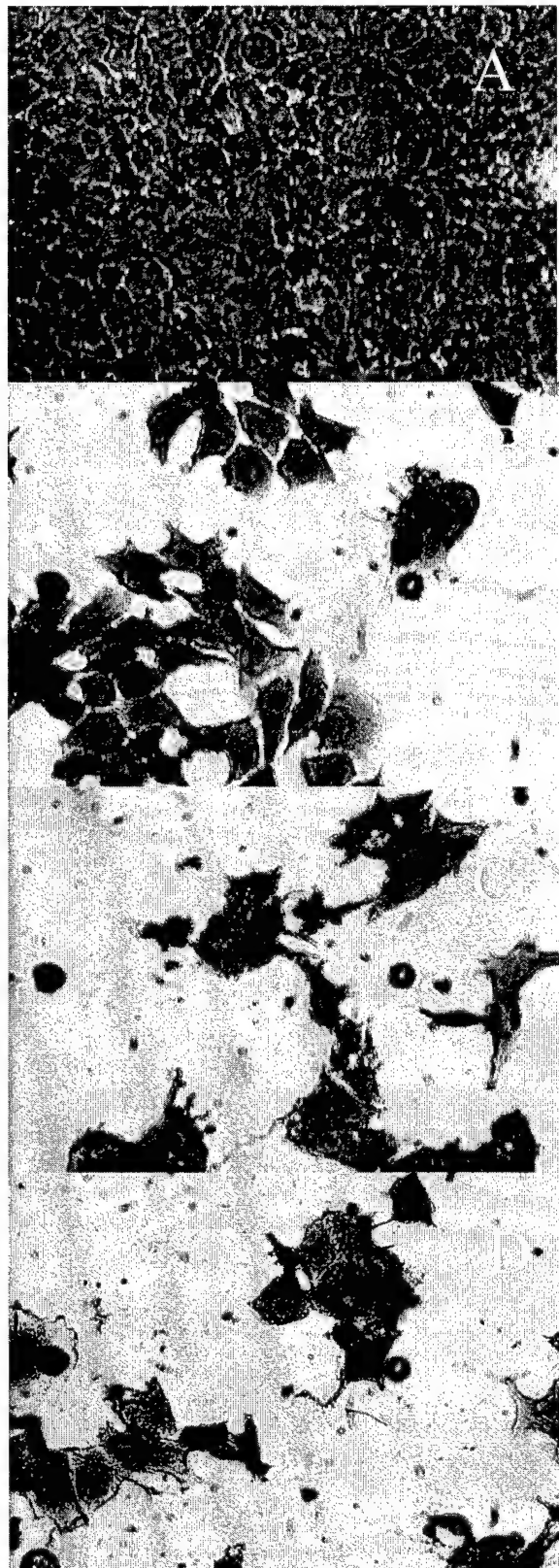


Fig. 3. Hypodense eosinophil line BJA.060.22 was co-cultured with MCF-7 tumor cells similarly to SD.031.22, however the ratios were 1:1 (B), 1:20 (C) and 1:40 (D) control tumor cells were cultured in media alone (A).

Fig. 4. Inhibition of MDA-MB-231 Tumor Cell Growth by Eosinophil Cell Lines

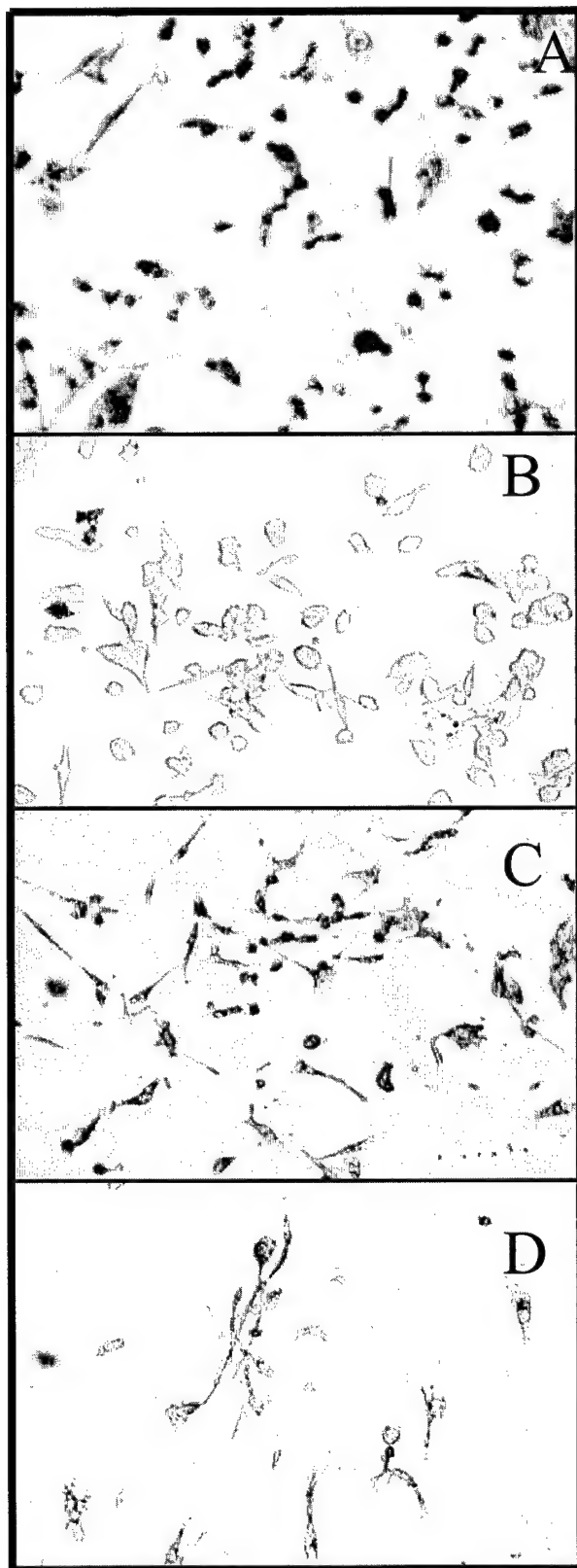


Fig. 4. Hyperdense BJA.060.24 was co-cultured with MDA-MB-231 tumor cells similarly to MCF-7 cells at E:T ratios 1:1 (B), 20:1 (C) and 40:1 (D). Control tumor cells were cultured in media alone (A).

**Fig. 5. Inhibition of MCF-7 Tumor Cell Growth
by Peripheral Blood Eosinophils**

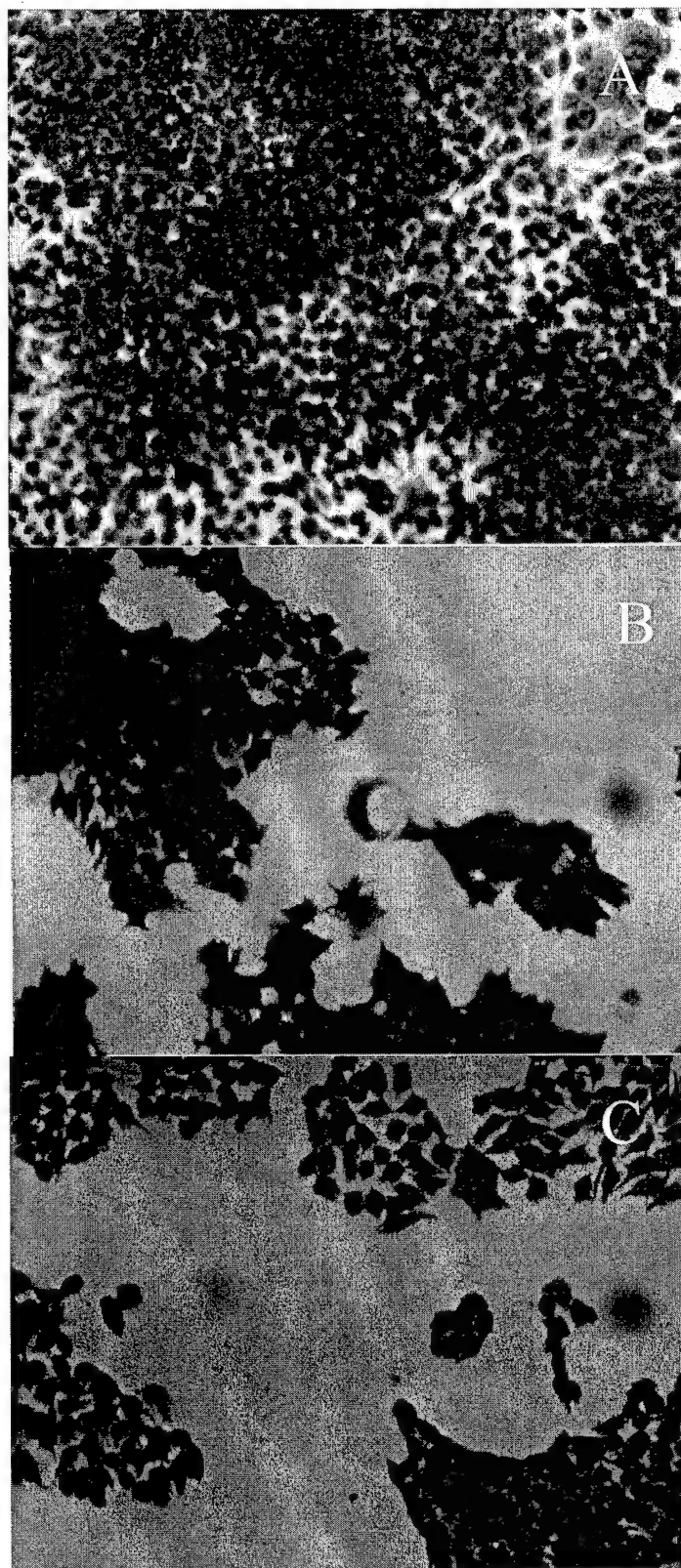


Fig. 5. Peripheral blood eosinophils, both hypodense (5B) and hyperdense (5C) show marked inhibition of cell growth when compared to that of the media control (5A) at 2:1 E:T ratios.

Fig. 6.

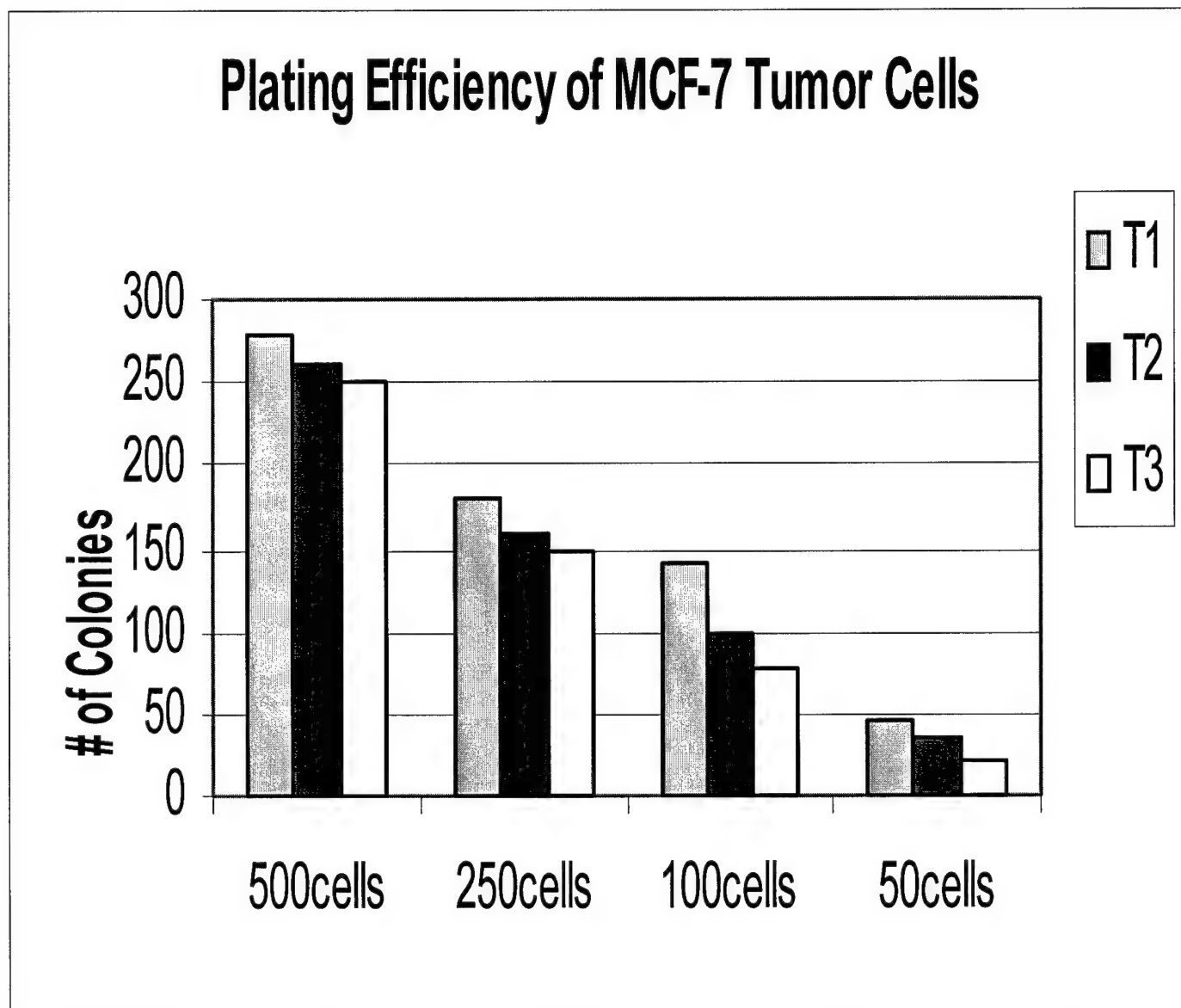


Fig. 6. MCF-7 cells were seeded into 6-well plates at 500, 250, 100 and 50 cells per well. The plates were then incubated for 10 days at 37°C, 5% CO₂. Plates were washed 3X with PBS, then stained with hematoxylin and eosin.

Fig. 7. Growth Inhibition of MCF-7 Tumor Cell Colony Formation by Peripheral Blood Hypodense and Hyperdense Eosionphils

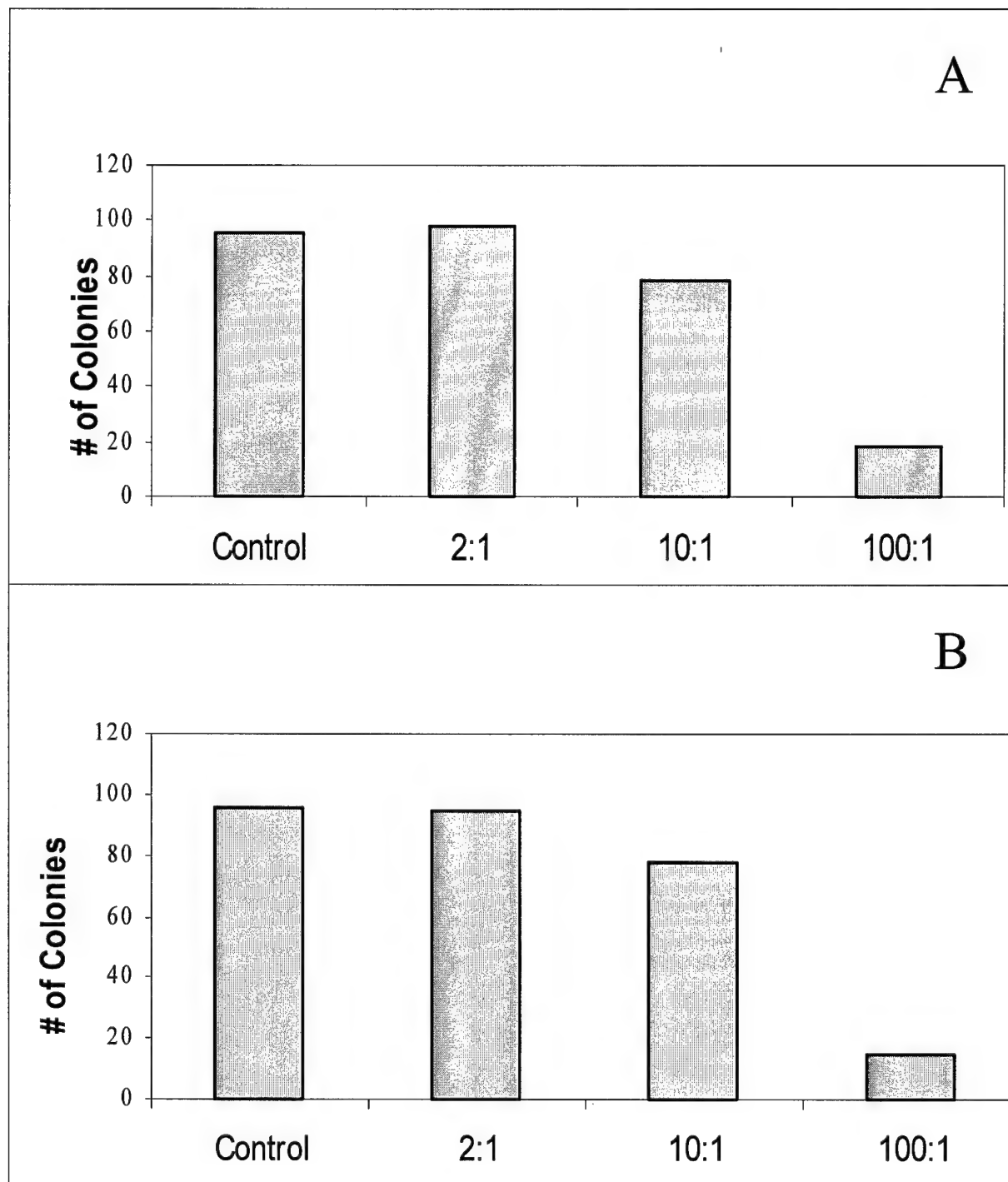


Fig. 7. MCF-7 cells were seeded into 6-well plates at 100 cells per well. 24hrs post seeding, hypodense (A) and hyperdense (B) eosinophils were added to the wells at E:T ratios of 2:1, 10:1 and 100:1. All plates were incubated for 10 days at 37°C, 5% CO₂.

Fig. 8. Growth Inhibition of MCF-7 Colony Formation by Eosinophil Cell Lines

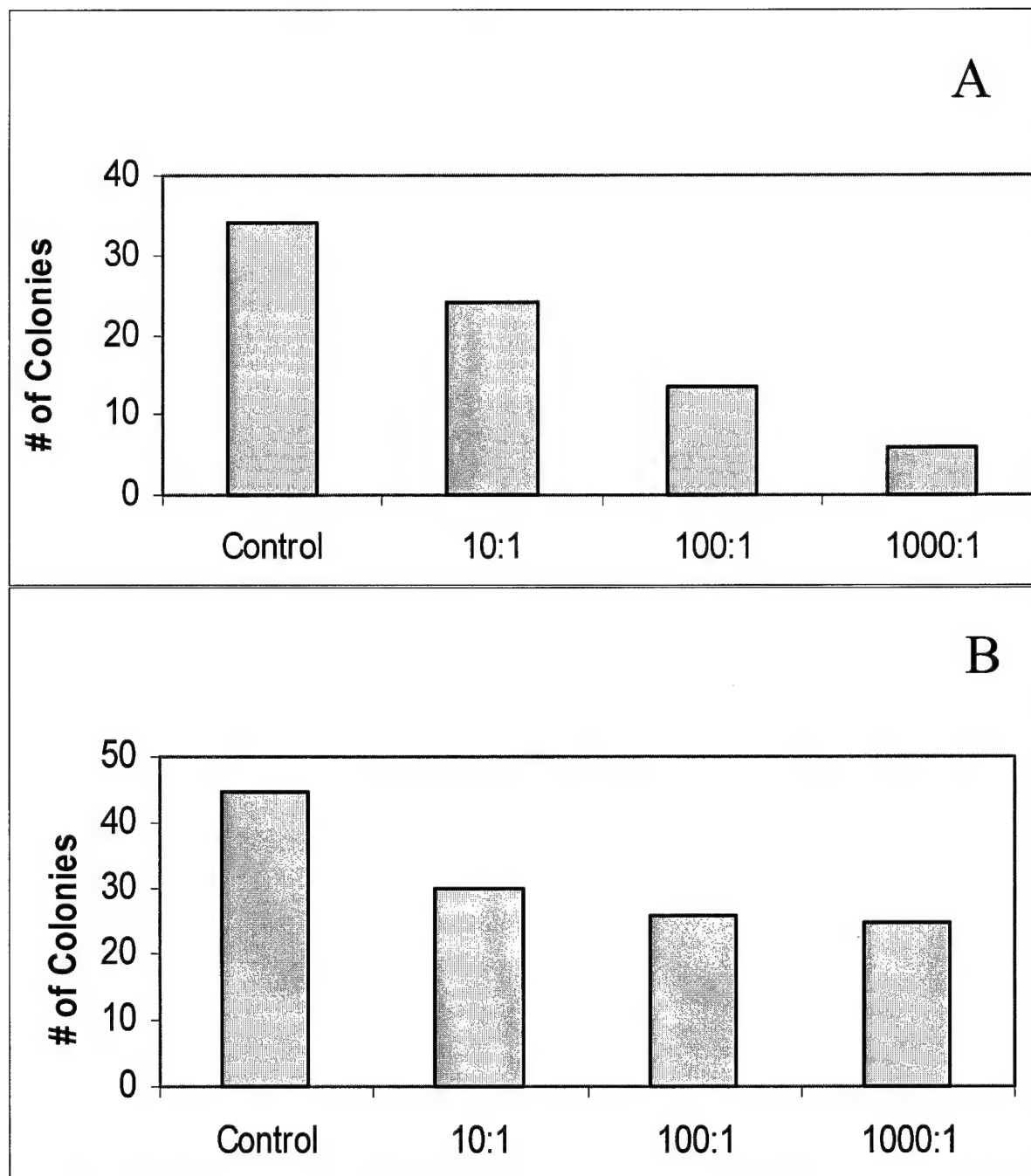


Fig. 8. BJA.060.22 (A) when cultured with MCF-7 cells at E:T ratios at 10:1, 100:1, and 1000:1 inhibited colony formation dose dependently, while BJA.060.24 (B) did not.

Fig. 9. Growth Inhibition of MDA-MB-231 Tumor Cell Colony Formation by Eosinophil Cell Lines

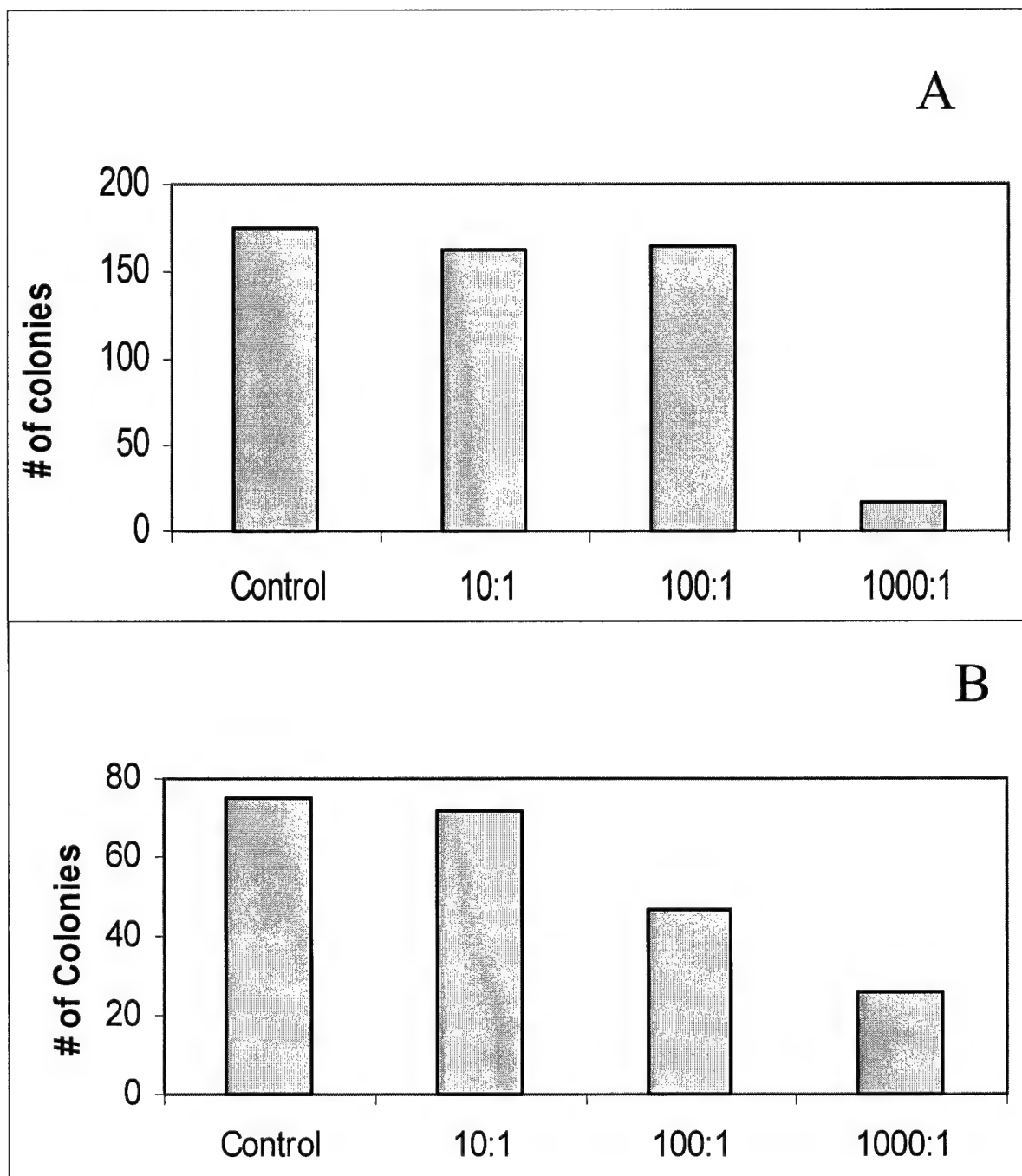


Fig. 9. BJA.060.24 (B) when cultured with MDA-MB-231 tumor cells at E:T ratios of 10:1, 100:1 and 1000:1 inhibited colony formation in a dose dependent manner, while BJA.060.24 markedly inhibited colony formation (93%) at the E:T ratio of 1000:1.

Table 1. CYTOKINE CONCENTRATIONS IN 24HR EOSINOPHIL CULTURE SUPERNATANTS (pg/ml)

Donor	IL-4		IL-5		TNF α		GM-CSF	
	22	24	22	24	22	24	22	24
1	>1000	>1000	440	435	50	63	0	0
2	316	3	0	0	100	56	0	0
3	>1000	631	0	0	50	16	0	0
4	>1000	0	nt	nt	129	200	nt	nt
5	200	20	0	0	100	224	nt	nt
6	8	>1000	0	186	10	7.9	450	450

Table 1. 24hr conditioned supernatants were tested for cytokines IL-4, IL-5, TNF α and GM-CSF using commercial enzyme linked immunoassay kits.

Fig. 10.

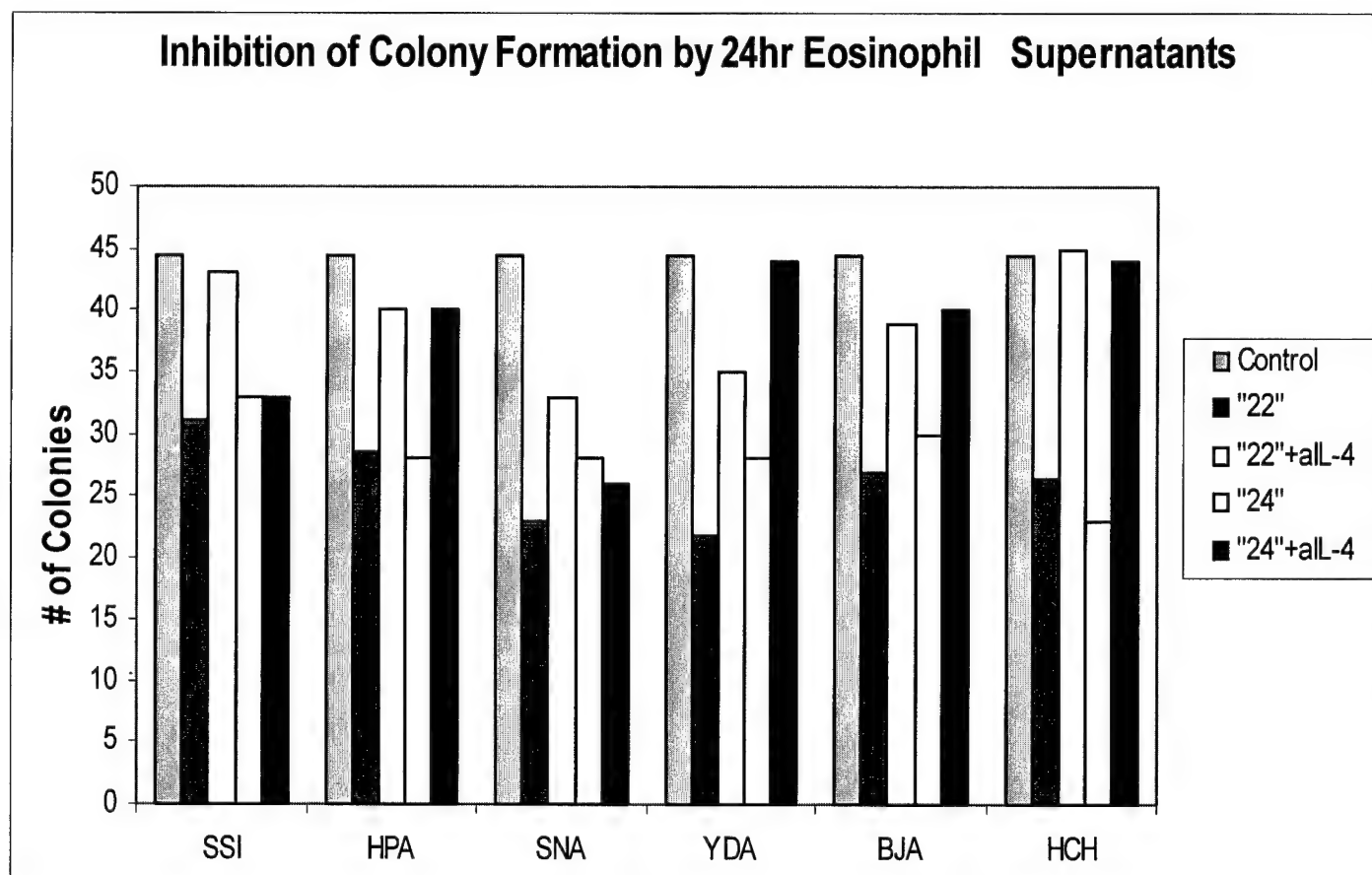


Fig. 10. MCF-7 tumor cells, (100 cells/well) with media, conditioned supernatants or with anti-IL-4 and supernatants for 10 days afterwhich the cells were washed 3X with PBS, then stained with hematoxylin and eosin.

Fig. 11. MCF-7 Multicellular Tumor Spheroids

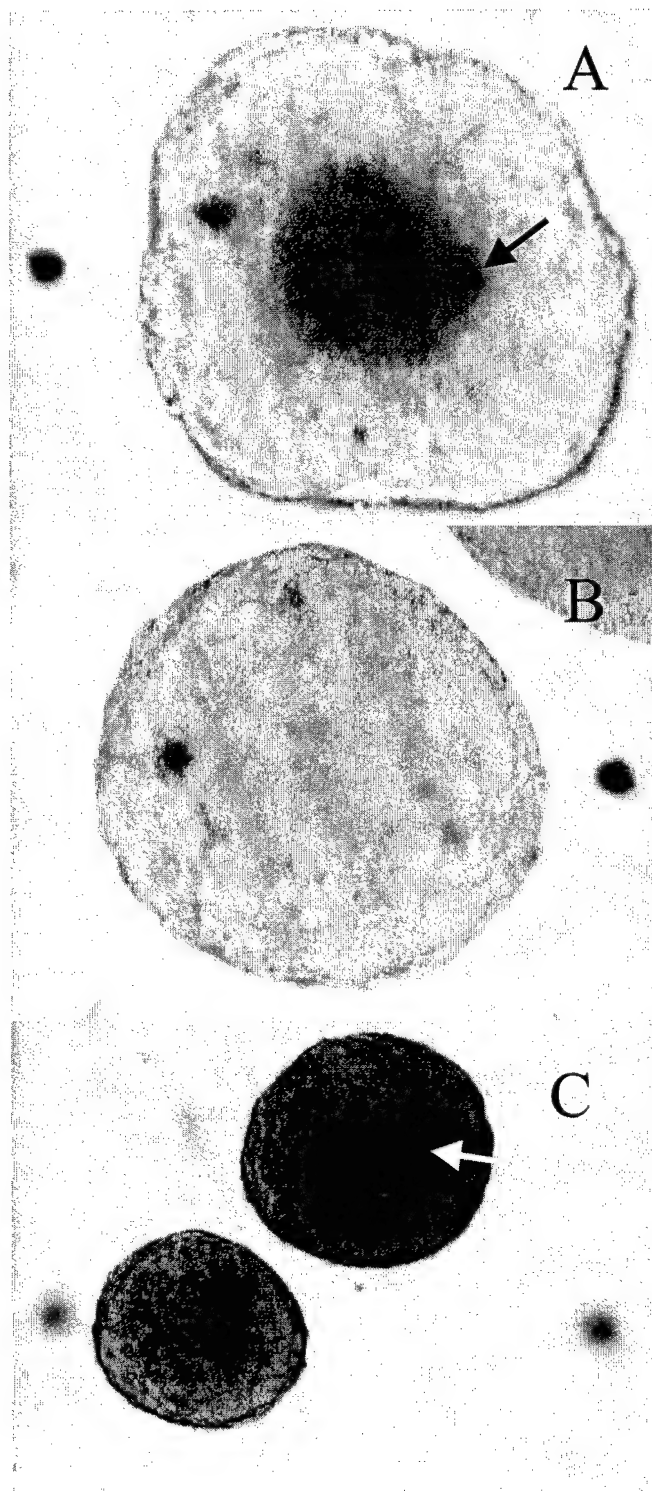


Fig. 11. MTS were developed by rocking 1×10^6 MCF-7 cells in a sealed T25 flask at 30 rpm for 48hr. At 37°C . The spheroids were then transferred to 100mm petri dishes containing 0.3% agar overlay, then cultured at 37°C , 5% CO_2 for 7-14 days.

Fig.12. Peripheral Blood Hypodense and Hyperdense Eosinophils Bind to MCF-7 Multi-cellular Tumor Spheroids

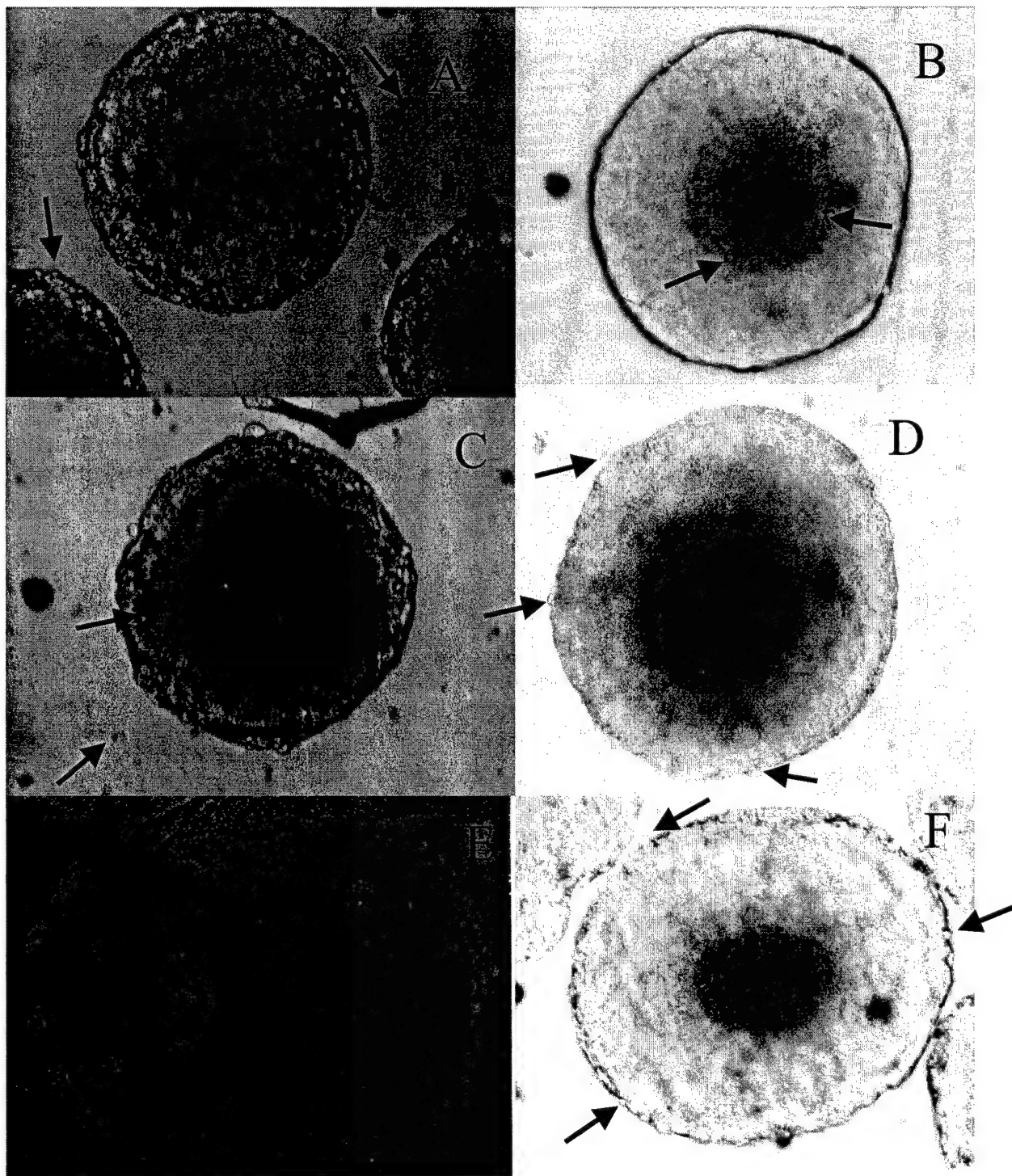


Fig. 12. Hypodense (A,C and E) and hyperdense (B,D and E) eosinophils from 3 individuals were cultured with 2 day-old MTS at an E:T ratio of 100:1 for 7 days. Arrows indicate bound eosinophils.

Fig. 13. Hypodense Eosinophilic Cell Line Binds to MCF-7 Multi-cellular Tumor Spheroids

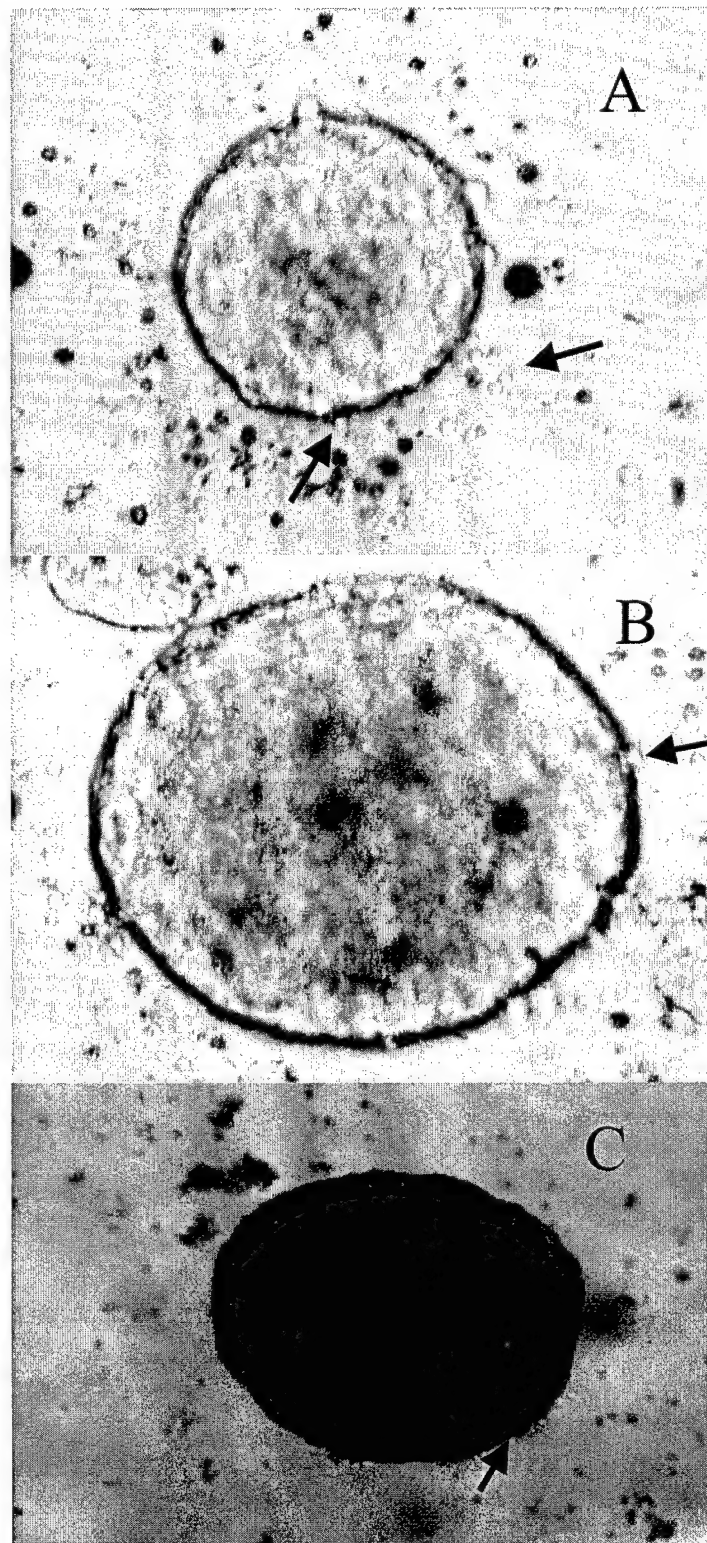


Fig. 13. Hypodense eosinophil cell line BJA.060.22 was cultured with 2-day old MCF-7 MTS at E:T ratios of 10:1 (A) 100:1 (B) and 1000:1 (C) for 7 days.

**Fig. 14. Eosinophil Infiltration of MCF-7
Multicellular Tumor Spheroid**



Fig. 14. Glutaraldehyde fixed tumor MTS:eosinophil complexes were prepared for transmission electron microscopic analysis. Eosinophils are seen in the MTS core (large arrow) as well within the spaces (small arrow).

AMERICAN
ASSOCIATION
FOR
CANCER RESEARCH

90th
ANNUAL MEETING

APRIL 10-14, 1999 • PHILADELPHIA, PA

Volume 40 • March 1999



In cooperation with



Proceedings

#3010 Hypodense and hyperdense eosinophils infiltrate MCF-7 breast multicellular tumor spheroids. Furbert-Harris PM, Harris D, Vaughn T, Parish-Gause D, Dunston GM, Abdelnaby A, Laniyan I, Oredipe O. *Howard Univ. Cancer Center, Wash., DC 20060.*

Our previous studies have indicated that activated eosinophils and eosinophilic cell lines inhibit *in vitro* growth of MCF-7 breast tumor cells. We have also shown that cytokines IL-4 and IL-5 also inhibit MCF-7 growth *in vitro*. In this study, MCF-7 multicellular tumor spheroids (MTS) were developed to study the effect of eosinophils, IL-4 and IL-5 on MTS growth. Hypo- and hyperdense metrizamide density gradient fractions of eosinophils from peripheral blood of individuals with mild to moderate eosinophilia were co-cultured with 2-day old MCF MTS in a medium containing bacto agar overlay in 5% CO₂ at 37C. Light microscopic analyses revealed attachment of effector cells to the spheroid borders. Moreover, the culture media was greater than 90% clear of effectors. At six days post co-culture, very large spheroids were observed in both test and control dishes; however the necrotic cores in the cocultures were more intense and larger than in the control. Furthermore, when MCF-7 tumor cells (1×10^6) were pretreated with IL-4 at 0.5 ng/ml, there was a dramatic decrease in the number of spheroids formed (90) compared to control (200). MTS are a more suitable model for *in vivo* development of micro metastatic nodules. These data strongly suggest that eosinophils are capable of infiltrating MTS and perhaps can release factors (cytokines like IL-4 and others which are capable of inhibiting tumor growth) into the extracellular spaces. These observations, along with the use of newly-established eosinophilic cell lines provide an exquisite model system for in depth studies of the role of eosinophils and cytokines in breast cancer and may offer potential therapeutic implications. D.O.D. #DAMD17-98-1-8107 and NIH# 5G12RR03048-12 RCMI Grant.

THE FASEB JOURNAL

A MULTIDISCIPLINARY RESOURCE FOR THE LIFE SCIENCES

Experimental Biology 99®
Washington, D. C.
April 17-21, 1999

ABSTRACTS PART I

Abstracts 1.1-486.12

Official Publication of the Federation of American Societies for Experimental Biology
Volume 13, Number 4, March 12, 1999

177.13

Eosinophilic Destruction of Breast Tumor Cells *In Vitro* is Mediated by Interleukin-4. Furbert-Harris PM, Anderson D, Parish-Gause D, Vaughn T, Brown R, Laniyan I, Dunston GM, Abdelnaby A and Oredipe O. Howard University, Washington, DC, 20060.

Previous studies in our laboratory have shown that activated eosinophils from individuals with eosinophilia will inhibit MCF-7 and MDA breast tumor cell growth *in vitro*. Moreover, eosinophilic cell lines established from these individuals inhibited MCF-7 colony formation in a dose dependent manner. In the present study, activated eosinophils were cultured for 24 hrs at 37C, 5% CO₂ and the conditioned media were captured for study. When MCF tumor cell monolayers were cultured with 24 hr supernatants, there was massive destruction of the monolayer similarly to that observed with the eosinophil-tumor co-cultures. MCF tumor cell colony formation was inhibited by both culture supernatants (37% and 31%, hypo- and hyperdense fractions, respectively) and by recombinant IL-4 (35%). Cytokine characterization of the supernatants by ELISA analysis indicated the presence of Interleukin-4 (IL-4) (> 1000ng/ml from both hypo- and normodense fractions) and TNF α (130 and 200 ng/ml from hypo- and hyperdense fractions, respectively). These results confirm the mouse data on eosinophil inhibition of tumor growth via IL-4 and strongly suggest a more active role for eosinophils as anticancer effectors. The regulation of this eosinophil anticancer activity by cytokines known to regulate eosinophils is under study. (Supported by DOD #DAMD17-98-1-8107 and NIH# 5G12RR03048-12 RCMI Grant)